

**23-ID-02****Committee:** Infectious Disease**Title:** Standardized Surveillance Case Definitions for Congenital Cytomegalovirus (cCMV) Infection and Disease

Check this box if this position statement is an update to an existing standardized surveillance case definition and include the most recent position statement number here: N/A.

**Synopsis:**

- This position statement creates standardized case definitions for cCMV infection and disease.
- Standardized case definitions for cCMV infection and disease are needed because multiple jurisdictions in the United States are conducting cCMV screening and surveillance activities but are using various methods and inclusion criteria for case ascertainment, reporting, and classification. As more jurisdictions pass legislation for newborn screening for cCMV, standardized case definitions for cCMV infection and disease can be used to understand the epidemiology of cCMV and compare trends across the United States.
- Case ascertainment criteria include laboratory criteria (the detection of CMV in neonatal urine, saliva, whole blood, or cerebrospinal fluid specimens, in amniotic fluid specimens, or umbilical cord or autopsy specimens), vital records criteria (infant death certificates), and healthcare records criteria (e.g., using ICD-10 diagnostic codes).
- Case classification criteria include clinical and laboratory criteria.
- Case classifications include confirmed cCMV infection, confirmed cCMV disease, and probable cCMV disease.

**I. Statement of the Problem**

Cytomegalovirus (CMV) infection during pregnancy can cause stillbirth, infant death, and a myriad of birth defects.<sup>1-3</sup> In the United States (U.S.), approximately 1 in 200 babies is born with congenital CMV (cCMV) infection; one out of 5 of these babies will present with clinical signs of cCMV disease in the neonatal period and/or have long-term health conditions.<sup>4</sup> cCMV is the most common infectious cause of developmental disabilities and non-genetic sensorineural hearing loss (SNHL) in U.S. children.<sup>5-8</sup> Nonetheless, the burden of cCMV disease is not fully understood.<sup>9-11</sup>

Surveillance of cCMV in the U.S. is complicated by several factors. First, most newborns with cCMV infection have no clinical signs at birth and, without universal cCMV screening, are not identified.<sup>12,13</sup> Second, neonatal clinical signs of cCMV disease are nonspecific and may be attributed to other conditions.<sup>14</sup> Third, postnatal CMV infection is common among infants, and a reliable diagnosis of cCMV infection or disease may not be possible unless specimens are collected within the first three weeks of life.<sup>15</sup> Finally, not all newborns with a laboratory diagnosis of cCMV infection receive a diagnostic code that would allow cases to be ascertained through a review of administrative data.<sup>16</sup>

**II. Background and Justification**

cCMV infection is responsible for an estimated 5-10% of cases of prelingual hearing loss among children less than 2 years of age, and an estimated 15-20% of moderate to profound bilateral SNHL among all U.S. children.<sup>7,17</sup> A substantial proportion of cCMV-related SNHL cases occur in children with cCMV infection who do not have apparent clinical signs at birth, including those who pass the newborn hearing screen.<sup>18</sup> Early identification and timely and appropriate intervention services are critical for improving developmental outcomes of deaf or hard-of-hearing children.<sup>19-22</sup> Consequently, the Joint Committee on Infant Hearing recommends that all infants who test positive for cCMV receive periodic audiologic monitoring beginning no later than three months of age to allow for the provision of appropriate amplification, early intervention, and family support.<sup>23</sup> Jurisdictional programs that monitor children with

cCMV infection or disease can assist providers and families by connecting them with resources and early intervention services. With current testing and surveillance practices, it is estimated that less than 10% of infants with cCMV infection are identified annually.<sup>24</sup>

In the U.S., cCMV infection has not yet been added by the Advisory Committee on Heritable Disorders in Newborns and Children to the Recommended Uniform Screening Panel.<sup>25</sup> In 2013, Utah began hearing-targeted screening for infants who refer on (i.e., do not pass) their newborn hearing screening and, in 2019, expanded screening to infants at high risk of cCMV disease, such as infants presenting with select clinical, laboratory, or brain imaging findings at birth, and those with maternal history of CMV infection during pregnancy.<sup>24</sup> In subsequent years, other states, including Connecticut, Florida, Iowa, Kentucky, Maine, New York, Pennsylvania, and Virginia, have passed legislation for hearing-targeted screening. In 2022, universal newborn cCMV screening was approved in Minnesota, and conditionally approved in New Jersey. Minnesota began screening newborns for cCMV using dried blood spot (DBS) specimens in February 2023.

While cCMV screening strategies and provider awareness may influence the likelihood of diagnosing cCMV infection or disease in infants, prevalence is also likely to vary across jurisdictions due to several factors, including maternal age and racial and ethnic composition of the population.<sup>4</sup> Currently, routine cCMV surveillance is conducted in 10 states. However, surveillance methods vary greatly, and a standardized cCMV case definition is lacking.<sup>24</sup>

Standardized case definitions for cCMV infection and disease would allow for comparisons of infection and disease prevalence among jurisdictions with different cCMV screening strategies and provide evidence to inform national newborn screening policy. Because most infants with cCMV infection never develop sequelae, quantifying the proportion of infants with cCMV disease likely to benefit from universal newborn screening, as compared to current standards of care (testing of newborns with clinical evidence of CMV-associated findings), is of critical importance.<sup>26</sup> Jurisdictions conducting or planning to implement cCMV surveillance with or without newborn screening may be uniquely suited to address this question. Jurisdictions may utilize data from cCMV surveillance to inform prevention efforts such as educational and awareness campaigns. Furthermore, standardized case definitions and validated methods for ascertaining cases of cCMV disease will be increasingly important to determine baseline case burden and prevalence of cCMV disease prior to licensure and routine use of CMV vaccines.<sup>27</sup>

This position statement establishes standardized cCMV case definitions for jurisdictions that wish to include cCMV infection and disease in their list of reportable conditions or conduct pilot studies or surveillance. Though this case definition is not intended for clinical use, public health officials could utilize it to provide feedback to healthcare providers and the public about the estimated burden and trends of cCMV infection and disease in their regions.

### **III. Statement of the Desired Actions to be Taken**

CSTE recommends the following actions:

1. Implement standardized surveillance case definitions for **cCMV infection and disease**.
  - A. Utilize standard sources (e.g., reporting\*) for case ascertainment for **cCMV infection and disease**. Surveillance for cCMV infection and disease should use the recommended sources of data to the extent of coverage presented in Section V.
  - B. Utilize standardized criteria for case ascertainment for **cCMV infection and disease** presented in Section VI and Table VI in Technical Supplement.
  - C. Utilize standardized criteria for case classification for **cCMV infection and disease** presented in Section VII and Table VII in Technical Supplement.

\* *Reporting: process of a healthcare provider, laboratory, or other entity submitting a report (case information) of a condition under public health surveillance to local, state, or territorial public health. Note: notification is the process of a local, state, or territorial public health authority submitting a report (case information) of a condition on the Nationally Notifiable Conditions List to CDC.*

#### **IV. Goals of Surveillance**

The goal of cCMV surveillance is to provide information on the burden of cCMV infection and disease in the U.S. and inform strategies to prevent cCMV-associated disability. cCMV surveillance may facilitate clinical follow-up, including connection to early intervention services for infants identified with cCMV infection and disease. Surveillance will help identify groups at higher risk of cCMV infection and disease and guide strategies to improve equity in access to services. Additionally, cCMV surveillance may provide data to inform provider and public awareness and educational activities.

#### **V. Recommended Data Sources and Methods for Surveillance**

Surveillance for cCMV infection and disease should use the following recommended sources of data and/or methodologies and the extent of coverage listed in Table V.

The most common data source for case ascertainment will be laboratory reporting. cCMV cases meeting the laboratory criteria as described in Section VI should be reported to the public health authorities within seven days of positive test result. Vital records data, birth defects registries, and diagnostic code data (see Appendix A) may be used as supplementary data sources for case finding. Universal newborn screening for cCMV, or targeted testing of high-risk populations, such as infants who refer on their newborn hearing screening, are used for identifying newborns with cCMV; follow-up testing may be required for infants who screen positive for cCMV. Laboratory testing for cCMV may be performed for diagnostic purposes when suspected by a clinician based on the presence of neonatal clinical signs, abnormal fetal ultrasound findings, or following a diagnosis of CMV infection in the gestational parent during pregnancy. Clinical information will be obtained from clinician reporting or review of medical records for case classification (see clinical requirements in Section VII).

**Table V. Recommended Sources of Data, Surveillance Methods, and Extent of Coverage for Ascertainment of Cases of cCMV Infection and Disease.**

<b>Source of Data/Methodology for Case Ascertainment</b>	<b>Coverage</b>	
	<b>Population-Wide</b>	<b>Sentinel Sites</b>
Clinician reporting	X	
Laboratory reporting	X	
Reporting by other entities, specify: <ul style="list-style-type: none"> <li>● Hospitals, clinics, or provider offices</li> <li>● Pharmacies</li> <li>● Other healthcare providers (e.g., midwives, public health nurses)</li> </ul>	X	
Death certificates	X	
Hospital discharge or outpatient records	X	
Data from electronic medical records	X	
Telephone or online survey		
School-based survey		
Other, specify: <ul style="list-style-type: none"> <li>● Autopsy reports</li> <li>● Vital Records</li> <li>● Birth Defect Registries</li> <li>● Early Hearing Detection and Intervention (EHDI) Information Systems</li> <li>● Early Intervention Referrals</li> </ul>	X	

## **VI. Criteria for Case Ascertainment**

Case ascertainment is the process through which public health identifies potential cases of a disease or condition using data reported or provided to public health by healthcare, laboratories, and other reporting entities. This public health reporting is triggered by the case ascertainment criteria (a single criterion or a combination of criteria) included in this position statement, and each initial report sent to public health should include common data elements and disease-specific data elements. Case ascertainment criteria are not intended to be used for clinical diagnosis purposes.

### **A. Narrative: A description of suggested criteria for case ascertainment of a specific condition and recommended reporting procedures.**

In public health jurisdictions conducting surveillance for cCMV, healthcare practitioners (including healthcare facilities and medical laboratories) should report cases of infection or disease that meet any of the laboratory, vital record, or healthcare record criteria for reporting, among children born in the jurisdiction. Case ascertainment criteria are intended to be used to identify potential cases for further investigation. Reported cases will be further reviewed for case classification.

Reporting of cCMV should be ongoing and routine.

### **Report to public health authorities infants or children meeting any of the following criteria:**

#### A1. Clinical Criteria for Reporting

N/A

#### A2. Laboratory Criteria for Reporting

- Detection of CMV DNA by polymerase chain reaction (PCR or other nucleic acid amplification testing [NAAT]) from infant<sup>†</sup> urine, saliva, whole blood (including dried blood spot [DBS]), or cerebrospinal fluid (CSF) specimen, **OR**
- Detection of CMV DNA by NAAT from an amniotic fluid specimen, **OR**
- Isolation of CMV in viral culture from infant<sup>†</sup> urine, saliva, whole blood, or CSF specimen, **OR**
- Isolation of CMV in viral culture from an amniotic fluid specimen, **OR**
- Demonstration of CMV antigen in a biopsy from umbilical cord or autopsy specimen by immunohistochemical methods (IHC), **OR**
- Detection of CMV antigen by antigenemia test in infant<sup>†</sup> whole blood specimen

<sup>†</sup> Infant is defined as less than one year of age. Note: Jurisdictions to determine reporting cutoff based on capacity, e.g., 45 days, 90 days, 6 months, one year.

#### A3. Epidemiologic Linkage Criteria for Reporting

N/A

#### A4. Vital Records Criteria for Reporting

- An infant aged one year or less whose death certificate lists cCMV or CMV as an underlying cause of death or significant condition contributing to death.

#### A5. Healthcare Record Criteria for Reporting

- A child aged 6 years or younger whose healthcare record contains a diagnosis\* of cCMV infection, **OR**
- An infant aged 45 days or younger whose healthcare record contains a diagnosis\* of CMV disease.

\* See Appendix 1 for diagnostic codes.

### **B. Disease-Specific Data Elements to be Included in the Initial Report**

Disease-specific data elements should be included in addition to the common data elements that are to be reported for all initial individual case reports (see CSTE Position Statement 09-SI-01 “Common Core Data Elements for

Case Reporting and Laboratory Result Reporting” <https://cdn.ymaws.com/www.cste.org/resource/resmgr/PS/09-SI-01.pdf>). Public health authorities do not expect that an initial report will contain all the information necessary for case investigation and case classification.

- Reason test was ordered for laboratory tests, when available.
- Date of death for death certificates.

## **VII. Case Definition for Case Classification**

This case definition for case classification is intended solely for public health surveillance purposes and does not recommend criteria for clinical diagnosis purposes. Once a public health agency has ascertained data on potential cases of a disease or condition from reporting entities, the public health agency assigns case statuses based on the case classifications included within this position statement.

### **A. Narrative: A description of criteria to determine how public health should classify a case of cCMV infection or disease.**

cCMV infection and disease are conditions caused by *in utero* infection with CMV. A wide spectrum of severity exists, from clinically inapparent infection to severe disease that is clinically apparent at birth or manifests as sequelae. The following guidelines are intended to be used for the purposes of cCMV surveillance and not intended to be used as a guide to clinical management of cCMV infection or disease.

#### **A1. Clinical Criteria**

Cases should be assessed according to absence or presence of clinical evidence as defined below and the clinical data should be included in the case investigation.

In the absence of a more likely alternative etiology:

- An infant with at least one of the following clinical signs during the neonatal period:<sup>28,29</sup>
  - Hepatomegaly
  - Splenomegaly
  - Petechial rash or purpura ("blueberry muffin rash"),

**OR**

- A child aged 6 years or younger with one or more of the following permanent conditions:<sup>28,29,30</sup>
  - Microcephaly (defined as head circumference measurement >2 standard deviations below the average (or <3rd percentile) for the same age and sex, notation in the medical record, or diagnostic code of microcephaly (e.g., ICD-10 code Q02),
  - Brain imaging abnormalities consistent with cCMV, such as intracranial calcifications, periventricular calcifications, leukomalacia, polymicrogyria, lissencephaly, pachygyria, schizencephaly, or ventriculomegaly
  - Sensorineural hearing loss
  - Seizures
  - Cerebral palsy
  - Chorioretinitis
  - Vision impairment, resulting from conditions consistent with cCMV, such as retinitis, retinal scarring, optic neuritis, optic atrophy, or brain damage resulting in cortical vision impairment

#### **A2. Laboratory Criteria\***

*Confirmatory Laboratory Evidence*<sup>†</sup>:

- Absence of a negative test (CMV DNA by NAAT or culture) on a urine specimen collected within 21 days of life, **AND**
- Detection of CMV DNA by NAAT from urine, whole blood (including dried blood spot [DBS]), or cerebrospinal fluid (CSF) collected from an infant within 21 days of life, **OR**
- Detection of CMV DNA by NAAT from amniotic fluid specimen, **OR**

- Isolation of CMV in viral culture from urine, whole blood, or CSF collected from an infant within 21 days of life, **OR**
- Isolation of CMV in viral culture from amniotic fluid specimen, **OR**
- Demonstration of CMV antigen in an autopsy specimen by IHC, **OR**
- Detection of CMV antigen by antigenemia test in whole blood collected from an infant within 21 days of life.

*Presumptive Laboratory Evidence:*

- Absence of a negative test (CMV DNA by NAAT or culture) on a urine specimen collected within 21 days of life, **AND**
- Detection of CMV DNA by NAAT from saliva collected from an infant within 42 days of life<sup>§</sup>, **OR**
- Isolation of CMV in viral culture from saliva collected from an infant within 42 days of life<sup>§</sup>, **OR**
- Detection of CMV DNA by NAAT from urine, whole blood, or CSF collected from an infant within 22–42 days of life<sup>¶</sup>, **OR**
- Isolation of CMV in viral culture from urine, whole blood, or CSF collected from an infant within 22–42 days of life<sup>¶</sup>.

*\* Note: The categorical labels used here to stratify laboratory evidence are intended to support the standardization of case classifications for public health surveillance. The categorical labels should not be used to interpret the utility or validity of any laboratory test methodology.*

<sup>†</sup> Only valid in the absence of a subsequent negative test on a urine specimen that was completed for confirmatory purposes.

<sup>§</sup> If CMV is detected in saliva, repeat testing should be performed using urine.

<sup>¶</sup> Only valid in the absence of a prior negative test on a urine specimen collected within 21 days of life.

### **A3. Epidemiologic Linkage Criteria**

N/A

### **A4. Case Classifications**

*Confirmed:*

- cCMV infection: Meets confirmatory laboratory evidence.
- cCMV disease: Meets clinical criteria AND confirmatory laboratory evidence.

*Probable:*

- cCMV disease: Meets clinical criteria AND presumptive laboratory evidence.

*Notes:*

- Cases of confirmed cCMV infection may be reclassified as confirmed cCMV disease if clinical evidence is subsequently identified after birth or later in childhood.
- Detection of CMV DNA by NAAT or culture from saliva collected from an infant within 21 days of life is considered as presumptive laboratory evidence because false-positive results may occur. Therefore, repeat testing using urine is recommended.
- Cases with clinical evidence of cCMV disease and presumptive lab evidence are classified as probable cCMV disease. This is done to reflect the uncertainty of lab evidence. Positive results on diagnostic testing performed after 21 days of life could pick up cases of postnatal CMV infection, which is often asymptomatic in term newborns but may present with clinical signs that may also occur in cCMV disease (e.g., hepatosplenomegaly, petechiae, thrombocytopenia), particularly in very low birth weight and preterm newborns.<sup>31</sup>
- Case classifications include confirmed cCMV infection to capture newborns that will mainly be identified via newborn screening, both universal and hearing-targeted. Most infected newborns will not have clinical signs of disease, including some who do not pass the newborn hearing screening but have normal hearing upon diagnostic audiologic evaluation. Whereas the estimated cCMV prevalence is 4.5 per 1,000 live births, incidence of acquired postnatal infection is at least 3% by 4-6 weeks of life<sup>32</sup>, increasing the probability of postnatal infection in infants with any positive test result in specimens collected between 22-42 days of life. Therefore, case classifications do not include “probable” cCMV infection, which could impact jurisdictional efforts on longitudinal data collection for permanent conditions.

## **B. Criteria to Distinguish a New Case of cCMV infection from Reports or Notifications which Should Not be Enumerated as a New Case for Surveillance**

A case should be enumerated as a new case if not previously reported.

Note: If a case was previously reported as cCMV infection but later meets criteria for cCMV disease the case would not be counted as a new case but a re-classification.

## **VIII. Period of Surveillance**

Surveillance is expected to be ongoing.

## **IX. Data Sharing/Release and Print Criteria**

CSTE recommends the following case statuses\* be included in the 'case' count released outside of the public health agency:

- Confirmed
- Probable
- Suspect
- Unknown

*\*Which case statuses are included in case counts constitute the "print criteria."*

Jurisdictions (e.g., States and Territories) conducting surveillance under this case definition can voluntarily submit de-identified case information to CDC, if requested and in a mutually agreed upon format.

Production of national data summaries and national data re-release for non-NNCs:

- Prior to release of national data summaries CDC should follow the CDC/ATSDR Policy on Releasing & Sharing Data, issued on April 16, 2003 and referenced in 11-SI-01 and custodians of such data should consult the CDC-CSTE Intergovernmental Data Release Guidelines Working Group report ([www.cste2.org/webpdfs/drgwgreport.pdf](http://www.cste2.org/webpdfs/drgwgreport.pdf)) which contains data release guidelines and procedures for CDC programs re-releasing state, local, or territorial-provided data.
- CDC programs have a responsibility, in collaboration with states, localities, and territories, to ensure that CDC program-specific data re-release procedures meet the needs of those responsible for protecting data in the states and territories.

## **X. Revision History**

Position Statement ID	Section of Document	Revision Description
23-ID-02	N/A	This is the first standardized surveillance position statement for cCMV infection and disease.

## **XI. References**

1. Iwasenko JM, Howard J, Ar buckle S, Graf N, Hall B, Craig ME, Rawlinson WD. Human cytomegalovirus infection is detected frequently in stillbirths and is associated with fetal thrombotic vasculopathy. *J Infect Dis* 2011; 203(11): 1526-1533.
2. Williams EJ, Embleton ND, Clark JE, Bythell M, Ward Platt MP, Berrington JE. Viral infections: contributions to late fetal death, stillbirth, and infant death. *J Pediatr* 2013; 163(2): 424-428.
3. Davis NL, King CC, Kourtis AP. Cytomegalovirus infection in pregnancy. *Birth Defects Res* 2017; 109(5): 336-346.
4. Fowler KB, Ross SA, Shimamura M, Ahmed A, Palmer AL, Michaels MG, Bernstein DI, Sanchez PJ, Feja KN, Stewart A, Boppana S. Racial and ethnic differences in the prevalence of congenital cytomegalovirus infection. *J Pediatr* 2018; 200: 196-201.
5. Boppana SB, Ross SA, Fowler KB. Congenital cytomegalovirus infection: clinical outcome. *Clin Infect Dis* 2013; 57(4):

- S178-S181.
6. Whitley RJ. Congenital cytomegalovirus infection: epidemiology and treatment. *Hot Topics in Infection and Immunity in Children. Adv Exp Med Biol* 2004; 549: 155-160.
  7. Satterfield-Nash A, Umrigar A, Lanzieri TM. Etiology of prelingual hearing loss in the universal newborn screening era: a scoping review. *Otolaryngol Head Neck Surg* 2020; 163(4): 662-670.
  8. Cannon MJ. Congenital cytomegalovirus (CMV) epidemiology and awareness. *Journal of Clinical Virology* 2009; 46(4): S6-S10.
  9. Tastad KJ, Schleiss MR, Lammert SM, Basta NE. Awareness of congenital cytomegalovirus and acceptance of maternal and newborn screening. *PLoS One* 2019; 14(8).
  10. Doutre SM, Barrett TS, Greenlee J, White KR. Losing ground: awareness of congenital cytomegalovirus in the United States. *Journal of Early Hearing Detection and Intervention* 2016; 1(2): 39-48.
  11. Pesch MH, Anderson C, Mowers E. Improving obstetric provider congenital cytomegalovirus knowledge and practices. *Infectious Diseases in Obstetrics and Gynecology* 2020.
  12. Dollard S, Grosse SD, Ross DS. New estimates of the prevalence of neurological and sensory sequelae and mortality associated with congenital cytomegalovirus infection. *Rev Med Virol* 2007; 17(5): 355-363.
  13. Ssentongo P, Hehnlly C, Birungi P, Roach MA, Spady J, Fronterre C, Wang M, Murray-Kolb LE, Al-Shaar L, Chinchilli VM, Broach JR, Ericson JE, Schiff SJ. Congenital cytomegalovirus infection burden and epidemiologic risk factors in countries with universal screening: a systematic review and meta-analysis. *JAMA Netw Open* 2021; 4(8).
  14. Ronchi A, Shimamura M, Malhotra PS, Sanchez PJ. Encouraging postnatal cytomegalovirus (CMV) screening: the time is NOW for universal screening! *Expert Rev Anti Infect Ther* 2017; 15(5): 417-419.
  15. Lazzarotto T, Guerra B, Lanari M, Gabrielle L, Landini MP. New advances in the diagnosis of congenital cytomegalovirus infection. *J Clin Virol* 2008; 41(3): 192-197.
  16. Campione A, Lanzieri TM, Ricotta E, Grosse SD, Kadri SS, Nussenblatt V, Prevots DR. Missing diagnoses of congenital cytomegalovirus infection in electronic health records for infants with laboratory-confirmed infection. *Curr Med Res Opin* 2022; 38(2): 273-275.
  17. Grosse SD, Ross DS, Dollard SC. Congenital cytomegalovirus (CMV) infection as a cause of permanent bilateral hearing loss: a quantitative assessment. *Journal of Clinical Virology* 2008; 41(2): 57-62.
  18. Fowler KB, McCollister FP, Sabo DL, Shoup AG, Owen KE, Woodruff JL, Cox E, Mohamed LS, Choo DI, Boppana SB. A targeted approach for congenital cytomegalovirus screening within newborn hearing screening. *Pediatrics* 2017; 139(2).
  19. Mayne AM, Yoshinaga-Itano C, Sedey AL, Carey A. Expressive vocabulary development of infants and toddlers who are deaf or hard of hearing. *Volta Rev* 1998; 100(5): 1-28.
  20. Kennedy CR, McCann DC, Campbell MJ, Law CM, Mullee M, Petrou S, Watkin P, Worsfold S, Yuen HM, Stevenson J. Language ability after early detection of permanent childhood hearing impairment. *N Engl J Med* 2006; 354(20): 2131-41.
  21. Watkin P, McCann D, Law C, Mullee M, Petrou S, Stevenson J, Worsfold S, Yuen HM, Kennedy C. Language ability in children with permanent hearing impairment: the influence of early management and family participation. *Pediatrics* 2007; 120(3): e694-701.
  22. Yoshinaga-Itano C, Sedey A, Coulter D, Mehl A. Language of early-and later-identified children with hearing loss. *Pediatrics* 1998; 102(5): 1161-1171.
  23. The Joint Committee on Infant Hearing. Year 2019 Position Statement: Principles and Guidelines for Early Hearing Detection and Intervention Programs. *JEHDI* 2019; 4(2): 1-44.
  24. Raines K, Nichols Heitman K, Leung J, Woodworth KR, Tong VT, Sugerman DE, Lanzieri TM. Congenital cytomegalovirus surveillance in the United States. *Birth Defects Research* 2022; 115(1): 11-20.
  25. HRSA. (2022, August). Recommended Uniform Screening Panel. <https://www.hrsa.gov/advisory-committees/heritable-disorders/rusp>
  26. Advisory Committee on Heritable Disorders in Infants and Children. (August 15, 2022) [Response letter to Dr. Megan Pesch's nomination of cCMV to the RUSP]. Retrieved from <https://www.hrsa.gov/sites/default/files/hrsa/advisory-committees/heritable-disorders/rusp/chair-letter-ccmv-nominators.pdf>
  27. Krause PR, Bialek SR, Boppana SB, Griffiths PD, Laughlin CA, Ljungman P, Mocarski ES, Pass RF, Read JS, Schleiss MR, Plotkin SA. Priorities for CMV vaccine development. *Vaccine* 2013; 32(1): 4-10.
  28. Cheeran MC, Lokensgard JR, Schleiss MR. Neuropathogenesis of congenital cytomegalovirus infection: disease mechanisms and prospects for intervention. *Clin Microbiol Rev* 2009; 22(1): 99-126.
  29. Swanson EC, Schleiss MR. Congenital cytomegalovirus infection: new prospects for prevention and therapy. *Pediatr Clin North Am* 2013; 60(2): 335-349.
  30. Goderis, J, De Leenheer, E, Smets, K, VanHoecke, H, Keymeulen, A, Dhooge, I. Hearing loss and congenital CMV infection: a systematic review. *Pediatrics* (2014) 134 (5): 972–982.
  31. Mukhopadhyay S, Meyer SA, Permar SR, Puopolo KM. Symptomatic postnatal cytomegalovirus testing among very low-birth-weight infants: indications and outcomes. *Am J Perinatol* 2016; 33(9): 894-902.
  32. Josephson, C, Caliendo, A, Easley, K, Knezevic, A, Shenvi, N, Hinkes, M, Patel, R, Hillyer, C, Roback, J. Blood transfusion and breast milk transmission of cytomegalovirus in very low-birth-weight infants: a prospective cohort study. *JAMA Pediatr* 2014 Nov;168(11):1054-62.



**XII. Coordination****Subject Matter Expert (SME) Consultants:****PRIMARY SME**

- (1) Tatiana M. Lanzieri, MD, MPH  
Medical Epidemiologist  
Viral Vaccine Preventable Diseases Branch  
Centers for Disease Control and Prevention  
(404) 639-3031  
[tmlanzieri@cdc.gov](mailto:tmlanzieri@cdc.gov)

**ADDITIONAL SMEs**

- (1) Kristen Nichols Heitman, MPH  
Epidemiologist  
Viral Vaccine Preventable Diseases Branch  
Centers for Disease Control and Prevention  
(404) 718-4670  
[knicholsheitman@cdc.gov](mailto:knicholsheitman@cdc.gov)
- (2) Suresh B. Boppana, MD  
Hugh Dillon MD Endowed Professor of Pediatrics  
Heersink School of Medicine  
University of Alabama at Birmingham  
(205) 638-2630  
[sbboppana@uabmc.edu](mailto:sbboppana@uabmc.edu)
- (3) Gail J. Demmler-Harrison MD  
Professor of Pediatrics  
Division of Infectious Diseases  
Baylor College of Medicine  
(832) 824-4327  
[gdemmler@bcm.edu](mailto:gdemmler@bcm.edu)
- (4) Karen B. Fowler, DrPH  
Professor of Pediatrics  
Heersink School of Medicine  
University of Alabama at Birmingham  
(205) 638-2549  
[karenfowler@uabmc.edu](mailto:karenfowler@uabmc.edu)
- (5) David W. Kimberlin, MD  
Professor of Pediatrics  
Co-Director, Division of Pediatric Infectious Diseases  
The University of Alabama at Birmingham  
(205) 638-2530  
[dkimberlin@uabmc.edu](mailto:dkimberlin@uabmc.edu)
- (6) Pablo J. Sanchez, MD  
Professor of Pediatrics  
Nationwide Children's Hospital  
The Ohio State University College of Medicine  
(614) 355-6638  
[pablo.sanchez@nationwidechildrens.org](mailto:pablo.sanchez@nationwidechildrens.org)
- (7) Mark R. Schleiss, MD  
American Legion and Auxiliary Heart Research  
Heart Research Foundation Professor of Pediatrics  
University of Minnesota Medical School  
(612) 626-9913  
[schleiss@umn.edu](mailto:schleiss@umn.edu)
- (8) Kate Russell Woodworth, MD, MPH, FAAP  
Medical Officer  
Infant Outcomes Monitoring  
Research, and Prevention Branch  
Centers for Disease Control and Prevention  
(404) 718-1178  
[Vnt0@cdc.gov](mailto:Vnt0@cdc.gov)

**Agencies for Response:**

- (1) Centers for Disease Control and Prevention  
Mandy K. Cohen, MD, MPH  
Director  
1600 Clifton Road NE  
Atlanta, GA 30329  
(404) 639-7000  
[Jbc7@cdc.gov](mailto:Jbc7@cdc.gov)

**Agencies for Information:**

N/A

**XIII. Author Information****Submitting and Presenting Author:**

- (1) Stephanie Browning McVicar, AuD, CCC-A  
EHDI Programs Manager  
Utah Department of Health and Human Services  
195 N 1950 W  
PO Box 144620  
Salt Lake City, UT, 84114-4620  
(801) 273-6600  
smcvicar@utah.gov

**Co-Authors:**

- (1) Active Member  
Max Sidesinger, MPH  
EHDI Epidemiologist/CMV Data Coordinator  
Utah Department of Health and Human Services  
195 N 1950 W  
PO Box 144620  
Salt Lake City, UT, 84114-4620  
(801) 273-4125  
msidesinger@utah.gov
- (2) Active Member  
Charla (Chas) DeBolt  
Nurse Consultant Advisor for Vaccine Preventable  
Diseases Surveillance  
Washington State Department of Health  
Center for Public Health Medical & Veterinary  
Science  
1610 NE 150th Street  
Shoreline, WA 98155  
(206) 418-5500  
Chas.DeBolt@DOH.WA.gov
- (3) Active Member  
Elizabeth Dufort, MD  
Medical Epidemiologist  
Minnesota Department of Health  
601 N Robert Street  
St. Paul, MN 55155  
(651) 201-5414  
elizabeth.dufort.contractor@state.mn.us
- (4) Active Member  
Tory Kaye, MPH  
Epidemiologist  
Minnesota Department of Health  
601 Robert Street N  
St. Paul, MN 55155  
(651) 201-5515  
tory.kaye@state.mn.us
- (5) Active Member  
Jessica Kumar, DO, MPH  
Medical Director, Bureau of Immunizations  
Division of Epidemiology, Center for Community  
Health  
New York State Department of Health  
ESP-Corning Tower, Room 651  
Albany, NY 12237  
(518) 473-4437  
jessica.kumar@health.ny.gov
- (6) Active Member  
Nicole D. Longcore, MPH  
Program Manager  
New York State Department of Health  
Division of Epidemiology  
Empire State Plaza, Corning Tower, Room 503  
Albany, NY 12237  
(518) 474-2875  
Nicole.Longcore@health.ny.gov
- (7) Associate Member  
Maryrose McInerney, PhD, CCC-A  
Associate Professor  
Montclair State University  
1515 Broad Street  
Bloomfield, NJ 07003  
(973) 655-3406  
mcinerneym@montclair.edu
- (8) Active Member  
Sondra D. Rosendahl, MS, LCGC  
Newborn Screening Follow-up Supervisor  
Minnesota Department of Health  
601 Robert Street N  
St. Paul, MN 55155  
(651) 201-5922  
sondra.rosendahl@state.mn.us

## Council of State and Territorial Epidemiologists Technical Supplement

**Table VI. Table of criteria to determine whether a case should be reported to public health authorities.**

Criterion	Reporting cCMV infection or disease
<i>Clinical Criteria for Reporting</i>	
N/A	
<i>Laboratory Criteria for Reporting</i>	
Detection of CMV DNA by NAAT from infant <sup>†</sup> urine, saliva, whole blood (including DBS), or CSF specimen	S
Detection of CMV DNA by NAAT from amniotic fluid specimen	S
Isolation of CMV in viral culture from infant <sup>†</sup> urine, saliva, whole blood, or CSF specimen	S
Isolation of CMV in viral culture from amniotic fluid specimen	S
Demonstration of CMV antigen in a biopsy from umbilical cord or autopsy specimen by IHC	S
Detection of CMV antigen by antigenemia test in infant <sup>†</sup> whole blood specimen	S
<i>Epidemiologic Linkage Criteria for Reporting</i>	
N/A	
<i>Vital Record Criteria for Reporting</i>	
An infant aged one year or less whose death certificate lists cCMV or CMV as an underlying cause of death or significant condition contributing to death.	S
<i>Healthcare Record Criteria for Reporting</i>	
A child aged 6 years or younger whose healthcare record contains a diagnosis* of cCMV infection	S
An infant aged 45 days or younger whose healthcare record contains a diagnosis* of CMV disease.	S

Notes:

S = This criterion alone is SUFFICIENT to report a case.

<sup>†</sup> Infant is defined as less than one year of age.

\*See Appendix 1 for diagnostic codes.

DBS, dried blood spot; CMV, cytomegalovirus; cCMV, congenital cytomegalovirus; CSF, cerebral spinal fluid; IHC, immunohistochemical methods; PCR, polymerase chain reaction (may also be called nucleic acid amplification test [NAAT])

**Table VII.A. Classification Table: Criteria for defining a case of cCMV infection or disease.**

Criterion	Case Classification			
	cCMV Infection	cCMV Disease		
	Confirmed	Confirmed	Probable	
<i>Clinical Evidence</i>				
Hepatomegaly		O		O
Splenomegaly		O		O
Petechial rash or purpura ("blueberry muffin rash")		O		O
Microcephaly <sup>††</sup>			O	O
Brain imaging abnormalities*			O	O
Sensorineural hearing loss			O	O
Seizures			O	O
Cerebral palsy			O	O
Chorioretinitis			O	O
Vision impairment <sup>¶</sup>			O	O
Absence of a more likely alternative etiology		N	N	N
Infant in neonatal period		N		N
Child aged 6 years or younger			N	N

<i>Laboratory Evidence</i>					
Absence of a negative test (CMV DNA by NAAT or culture) on a urine specimen collected within 21 days of life	N	N	N	N	N
Detection of CMV DNA by NAAT from urine, whole blood (including DBS), or CSF collected within 21 days of life	O	O	O		
Detection of CMV DNA by NAAT from amniotic fluid specimen	O	O	O		
Isolation of CMV in viral culture from urine, whole blood, or CSF collected within 21 days of life	O	O	O		
Isolation of CMV in viral culture from amniotic fluid specimen	O	O	O		
Demonstration of CMV antigen in an autopsy specimen by IHC	O	O	O		
Detection of CMV antigen by antigenemia test in whole blood collected within 21 days of life	O	O	O		
Detection of CMV DNA by NAAT from saliva collected within 42 days of life <sup>§</sup>				O	O
Isolation of CMV in viral culture from saliva collected within 42 days of life <sup>§</sup>				O	O
Detection of CMV DNA by NAAT from urine, whole blood or CSF collected at 22–42 days of life				O	O
Isolation of CMV in viral culture from urine, whole blood, or CSF collected within 22–42 days of life				O	O
<i>Epidemiologic Linkage Evidence</i>					
N/A					

**Notes:**

N = All “N” criteria in the same column are NECESSARY to classify a case.

O = At least one of these “O” (ONE OR MORE) criteria in each category (categories=clinical evidence, laboratory evidence, and epidemiologic evidence) in the same column—in conjunction with all “N” criteria in the same column—is required to classify a case.

<sup>††</sup> Microcephaly is defined as head circumference measurement >2 standard deviations below the average (or <3rd percentile) for the same age and sex, notation in the medical record, or diagnostic code of microcephaly (e.g., ICD-10 code Q02).

\* Brain imaging abnormalities consistent with cCMV, such as intracranial calcifications, periventricular calcifications, leukomalacia, polymicrogyria, lissencephaly, pachygyria, schizencephaly, ventriculomegaly.

<sup>†</sup> Vision impairments resulting from conditions consistent with cCMV, such as retinitis, retinal scarring, optic neuritis, optic atrophy, or brain damage resulting in cortical vision impairment.

<sup>§</sup> If CMV is detected in saliva, repeat testing should be performed using urine.

DBS, dried blood spot; CMV, cytomegalovirus; cCMV, congenital cytomegalovirus; CSF, cerebral spinal fluid; IHC, immunohistochemical methods; NAAT, nucleic acid amplification test

**Table VII.B. Classification Table: Criteria to distinguish a new case of cCMV infection from reports or notifications which should not be enumerated as a new case for surveillance.**

Criterion	cCMV Infection	cCMV Disease	
	Confirmed	Confirmed	Probable
<i>Criteria to distinguish a new case</i>			
A case should be enumerated as a new case if not previously reported.	S	S	S

**Appendix 1. Diagnostic codes for case ascertainment of congenital cytomegalovirus (cCMV) infection for surveillance purposes.****Diagnostic codes for Congenital Cytomegalovirus Infection and Cytomegalovirus Disease**

ICD-10-CM P35.1	Congenital cytomegalovirus infection
ICD-10-CM B25.x	Cytomegaloviral disease

**Abbreviations:** ICD-10-CM, International Classification of Disease, 10th Revision

**Potential Data Sources.** Potential data sources for using ICD-10 (or future versions) diagnostic codes for ascertainment of cCMV cases for surveillance purposes include, but are not limited to, birth and death certificates, hospital inpatient, outpatient, or discharge records, and data from electronic medical records.

**Considerations:** Laboratory confirmation of cCMV should be performed on specimens collected within the first 21 days of life. If specimens are collected after 21 days, then it is challenging to distinguish congenital from postnatal infection. Researchers that have used administrative, hospital discharge, or electronic medical record data have typically used an interval of 30-90 days to ascertain potential cases of cCMV. Restricting case-finding to those infants with diagnostic codes within a certain period after birth will help to minimize the likelihood of ascertainment of postnatal CMV cases and allows for delays in conducting testing, submitting claims, or reporting results. In the absence of reporting of laboratory test results, case ascertainment using diagnostic codes may assist with cCMV surveillance and provide data to examine trends over time, although caution is recommended for interpreting findings due to several limitations outlined below.

**Limitations:** To date, the validity of diagnostic codes for cCMV surveillance has not been evaluated. Coding errors, under- or over-reporting, and inclusion of postnatal cases are possible. One study found that only 1 in 10 infants with laboratory-confirmed CMV infection within 21 days of life had a diagnostic code for cCMV.<sup>16</sup> The extent to which the codes for congenital cytomegalovirus infection (ICD-10-CM code P35.1) and cytomegalovirus disease (ICD-10-CM code B25.x) are used interchangeably for cCMV disease during infancy is unknown. Unpublished analyses of administrative data using both conditions to identify cases among infants within 45 days of life indicate that 53-56% of infants have a cCMV infection code (P35.1) alone, 18-25% have a CMV disease code (B25.x) alone, and 21-28% have both codes.